Research Article

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# Genome-wide Association Study using the Canine SNP20 Beadchip for Canine Hip Dysplasia in Labrador Retriever

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#### **Abstract**

Many dog genetic disease, which can mean that the rate of inbreeding is the result. Canine hip dysplasia (CHD) is a common hereditary developmental disease of the coxofemoral joints and one of the most important bone and joint diseases. CHD is characterized by subluxation of the femoral head and deformation of the acetabulum leading to a painful osteoarthrosis. The radiographic diagnosis is sometime possible when the disease has markedly progressed. Looking up now studies with dog genetic disease a dwarfism (Germany shepherd), progressive retinal atrophy (Mastiff), malignant fever (all variety). With molecular biological techniques are being developed. This study, the hip dysplasia and early diagnosis that can be genetic markers to develop a more 22K SNP chip using the hip dysplasia in the X-ray diagnostics have Labrador retriever 48 of blood from the Wizard Genomic DNA Purification Kit using the DNA was extracted. This consists of SNP information, the entire 22,362 Canine SNP22 Beadchip used, SNP genotyping was carried out. Associated with hip phenotype for QTL detection for each SNP marker regression analysis using R statistical program (single SNP regression analysis) through the expression of hip additive effect of SNP marker for the evaluation. As the primary analysis of hip dysplasia in SNPs they have significance to the 4, 38, X chromosome respectively 11, 4, 10 results were excavated a total of 25 (genome-wise *P* < 0.001).

Keywords: Genome-wide association study, Single nucleotide polymorphism, Canine, hip dysplasia, Marker

## Introduction

The dog is attractive species to discover genes underlying the homologous, relevant diseases in humans due to the high relevance of canine diseases to those in humans and the intrinsic importance of dogs to humans as special companions with shared environments. The domestic dog species is divided into over 300 pure breeding populations known as breeds. Many breeds are characterized by reduced genetic diversity related to small numbers of founders, popular sires whose allelic pool is over represented in subsequent generations, and changes in breed popularity over time (Quignon *et al.*, 2007). Shared phenotypic characteristics of human and canine hip dysplasia (HD) are hip joint laxity accompanied by hip subluxation. Human HD, referred to as developmental dysplasia of the hip, occurs with a frequency ranging from 5, 13% (Zhou *et al.*, 2010). Canine HD, a complex trait, is a major veterinary problem

occurring with a frequency up to 75% in mixed and pure breed dogs of the approximately 70 million dogs in American households. Canine HD is a complex genetic skeletal disease observed in the majority of breeds of dogs and in almost all large-sized breeds of dogs. Canine HD is characterized by delayed onset of the capital femoral ossification, laxity of the joint, loss of congruency between acetabulum and femoral head, subluxation up to luxation, and often secondary osteoarthrotic changes. The first report of canine HD was reported in 1935. A study in 2003 showed that the prevalence of Canine HD was 19.3% in the general population of pet dogs. The percentage of dysplasia for these breeds in that study was 35.4% for Rottweiler's, 32.9% for German Shepherds, 30.3% for Golden Retrievers and 27% for Labrador Retrievers (Nganvongpanit *et al.*, 2008). The Labrador retriever demonstrates a 3.4-fold increase in risk for the development of HD (Goldberg and Rubin, 1989). Moreover, the majority of canine HD (80%) had osteoarthritis (OA). So far, the initiating factors are unknown, and the rate and extent of the development of HD are variable, but the risk factors are both genetic and environmental. As mentioned above, most of the dogs with HD in the above mentioned study also had OA, but the standardized diagnostic protocol consists of the clinical sign, a physical examination and evaluation of the radiographic results, which cannot detect OA in its early stages (Nganvongpanit *et al.*, 2008).

Worldwide, the Orthopedic Foundation for Animals (OFA) has been scoring hip radiographs and releasing some of the records publicly over the last 40 years. In a previous study showed that a consistent genetic improvement has accumulated. The genetic improvement was limited by the fact that the selection criteria of the majority of the breeding dogs had low accuracy. Criteria for canine HD evaluated in scoring schemes include congruency of acetabulum and femoral head, the extent to which the femoral head is fully encompassed by the acetabulum, shallowness of the acetabulum and (sub-)luxation of the hip joint, and smoothness and radiographic sharpness of bony structures. In Europe, Fédération Cynologique Internationale (FCI) has made uniform as much as possible the scoring of hip joints in all registered breeds of dogs. Frequency and distribution of severity of scores vary largely between breeds, but clinical and radiographic signs are consistent across the different breeds of dogs (Marschall and Distl, 2007).

In the past, the uses of molecular genetics in commercial applications of marker-assisted selection have focused on the use of individual genes or a few quantitative trait loci (QTL) linked to markers. But until recently single-nucleotide polymorphism (SNP) genotyping technologies have provided for the first time the opportunity to rapidly screen large numbers of samples for genetic variation associated with increased risk for disease. SNP Microarray provides data on both genotype and signal intensity, the combination of which can be used to generate information on chromosomal segment copy number.

Previous study is canine HD in German shepherd dogs the using micro satellites (MS) marker. A whole genome scan for QTL was performed in German shephered dogs and 11 paternal half sib families, including a total of 459 purebreed German shepherd dogs with sires, dams, and offspring, were genotyped for 261 microsatellites. QTL for canine HD were located on nine different canine chromosomes: 1, 3, 4, 8, 9, 16, 19, 26, and 33. (Marschall and Distl, 2007).

We present data for the first time to demonstrate that canine HD is predictable from genomic data so that selection decisions. This implies that human HD could also be predicted and suitable preventative management could be applied to identify susceptible individuals who may be missed by physical screening and ultrasound and reduce the prevalence hip OA by pre-emptive intervention.

#### **Materials and Methods**

#### **Animal**

Forty eight native Labrador Retriever dogs were categorized into two groups. Twenty four dogs were low risk hip dysplasia (HD) group consisting of 13 male and 11 female. Another group had 24 dogs which were high risk HD group consisting of 13 male and 11 female. Blood samples were collected for genomic DNA isolation using Wizard genomic DNA purification kit (Promega, USA) from 48 and analyzed concentration and purity at absorbance of 260 nm and 280 nm using ND-1000 spectrophotometer (Nanodrop, USA).

#### Phenotypic traits

Radiology has commonly been used to diagnose CHD. The technique has been standardized worldwide, although there is some variation in radiograph evaluation. There are three (somewhat different) international scoring methods: the FCI, the OFA, and the British Veterinary Association/Kennel Club methods. The FCI scoring method is used in most mainland European countries, Russia, South America, and Asia. The OFA approach is used exclusively in the USA and Canada, and the BVA/KC method is used in Britain, Ireland, Australia and New Zealand. We used the OFA method, OFA method divided 7group and each group is excellent (0 point), good (1~6point), fair (7~8 point), borderline (9~12), mile (13~18), moderate and severe (18~). This study radiograph score divided into two groups low risk, high risk was divided into. Low risk group average is 3.3 point; high risk group average is 15.3 point.

# **SNP Genotyping**

Canine sample genotyped on the infinium canine SNP22 beadChip (Illumina Inc., San Diego, CA) with ~22,000 SNPs across he genome (http://www.illumina.com/documents/ products/datasheets/datasheet\_canine\_snp20.pdf). The majority (92.3 %) of the Illumina SNPs had completed calls. We also removed SNPs with minor allele frequency (MAF) below 1%. The final analysis contained 21,455 SNPs for the Illumina array, the mean and median MAF were 0.2589 and 0.2399, respectively.

#### Statistical Analysis

Genotypes were tested for Hardy-Weinberg equilibrium (HWE) to identify possible genotyping errors using the Chi-square test in the R/SNPassoc Package (R Development Core Team). SNPs with HWE (P < 0.05), fail to call (> 80%), monomorphic SNPs and minor allele frequency (< 0.01) were removed in this

QTL study. The association between a given SNP and the outcome was assessed using the R/SNPassoc package (http://cran.r-project.org/web/packages/SNPassoc/index.html) (model 1).

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[model I]
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 $\mathbf{y} = \boldsymbol{\mu} + \mathbf{Sex_i} + \mathbf{SNP_j} + \mathbf{e_{ij}}$ 

y: hip dysplasia phenotype

μ: Full average

Sex.: Genotype effect (i=Female, Male and Male Castration)

SNP: Genotype effect (j=AA, AB, BB)

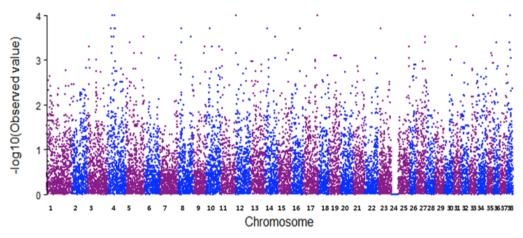
 $e_{ij}$ : Random error, N~(0,  $\sigma_{\epsilon}^2$ )

### Results

SNP analysis identified three different chromosomes for canine HD and each chromosome location (Fig. 1-2, Table 1.). Figure 1 and Table 1 show the plots summarize the genome-wide association results for hip dysplasia. The genome-wide P value ( $\log_{10}$  p) of the t test for the SNP effect are plotted against position on 4, 38, X chromosome and Locate on 4, 38, X chromosome SNPs position, t-statistical analysis information. Chromosome number is plotted in the x-axis. Horizontal line indicates the threshold P < 0.0001.

**Table 1.** A significant level of association canine hip dysplasia statistical analysis of SNP.

CHR	SNP	Position	Estimations	t-statistics	Permutation_P	log(P)
4	BICF2P582065	21279946	8.544	4.507	0.0002	3.69897
	BICF2S23632496	23272314	<b>7.</b> 51	4.167	0.0007	3.15496
	BICF2P95705	24055834	6.066	3.746	0.0003	3.52288
	BICF2P750308	24066370	6.066	3.746	0.0003	3.52288
	BICF2P783744	27241295	6.453	3.834	0.0005	3.30103
	BICF2P331356	27447828	11.3	4.608	0.0001	4.00004
	BICF2P1188097	27555277	6.257	3.748	0.0001	4.00004
	BICF2P676865	33248207	9.49	4.262	0.0003	3.52288
	BICF2S23030416	38226314	6.925	4.012	0.0001	4.00004
	BICF2G630166744	38258702	6.664	3.957	0.0002	3.69897
	BICF2S23110118	42752780	7.034	4.059	0.0002	3.69897
38	BICF2S2373811	16185735	-7.247	-3.067	0.0004	3.39794
	BICF2P817345	16696343	5.764	3.895	0.0001	4.00004
	BICF2G63073656	16775306	5.025	3.415	0.0006	3.22192
	BICF2S23036049	18480476	4.989	3.274	0.0009	3.04581
X	BICF2S2297550	91191649	6.197	3.527	0.0009	3.04581
	BICF2S22933746	99354996	7.311	4.011	0.0002	3.69897
	BICF2G6306331	108201633	7.5	3.623	0.0002	3.69897
	BICF2P227876	109116340	7.5	3.623	0.0002	3.69897
	BICF2G6305980	109652960	6.889	3.582	0.0005	3.30103
	BICF2P162522	111424321	7.367	4.106	0.0001	4.00004
	BICF2P593928	111997007	7.367	4.106	0.0001	4.00004
	BICF2S23044648	121338961	-8.175	-3.686	0.0005	3.30103
	BICF2P1430795	121460633	-8.175	-3.686	0.0005	3.30103
	BICF2S23222536	121643766	8.175	3.686	0.0005	3.30103



**Figure 1.** Manhattan plot of P-value for HIP score CHD in LR. The log inverse P-values estimated for each polymorphism are plotted in the y-axis.

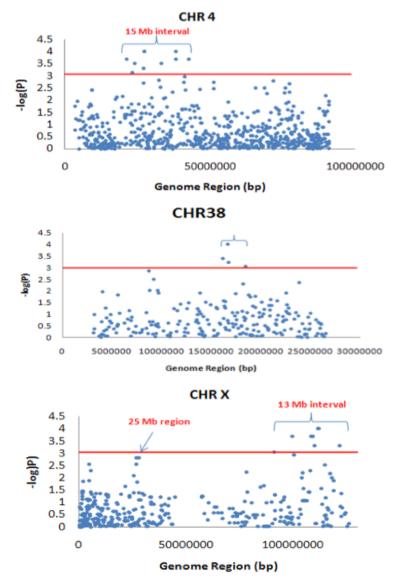


Figure 2. Manhattan plots; 4, 38, X chromosome associated hip dysplasia on the location of SNP.

Table 2 shows gene function and notable nearby gene including significant SNPs. as shows notable nearby gene function is divided 4groups. Each group is cells and cell binding, growth and immune, signal transduction, formation of institutions. Notable nearby gene is caitenin alpha 3 (CTNNA3), stork head BOX1 (STOX1), nucleolar protein GU2 isoform 2 (DDX50), procollagen type XIII alpha1 (COL1A1), ubiauitin specific protease54 (USP54), heparin sulfate N-deacetylase (Ndst), urokinase plasminogen activator preproprotein (PLAU), voltage dependent anion channel 2 (VDAC2), SH2 domain containing 4B (Sh2d4b), neurequlin 3 (NRG1), BMP 3b precursor, USP6 N-terminal like protein (USP6NL), ER-Golgi intemediate compartment 32kDa prote, Dual specificity protein phosphatase1, RIKEN Cdna 4930513F16, Transforming growth factor (TGF), lysophospholopase - like1, MOCO Sulphurase c-terminal domain containing (MOSC), leucine rich repeats and calponin homology (LRCH), glutamine ammonia ligase (GLUL), odd oz/tenm homolog 1 (ODZ1), hypoxantine phosphoribosyltransferse (HGPT), DEADLH box polypeptide2, solute carrier family 9 (SLC9A9), zinczinc finger protein of the cerellum3, fibroblast growth factor 13 isoform4 (FGF13 isoform4), ATP binding cassette (Abc) as notable nearby gene revealed a total of 27.

**Table 2.** 4, 38, X chromosome associated SNP function and notable nearby gene.

CHR	SNP	Notable Nearby Gene	Function
4	BICF2P582065	caitenin alpha 3	cells and cell binding
	BICF2S23632496	storkhead BOX1, nucleolar protein GU2 isoform 2	-
	BICF2P95705	procollagen type X III alpha1	cells and cell binding
	BICF2P750308	ubiauitin specific protease54	growth and immune
	BICF2P783744	ubiauitin specific protease54	growth and immune
	BICF2P331356	heparin sulfate N-deacetylase	formation of institutions
	BICF2P1188097	urokinase plasminogen activator preproprotein, voltage dependent anion channel 2	signal transduction
	BICF2P676865	SH2 domain containing 4B, neurequlin 3	growth and immune
	BICF2S23030416	BMP 3b precursor, USP6 N-terminal like protein	growth and immune
	BICF2G630166744	BMP 3b precursor, USP6 N-terminal like protein	growth and immune
	BICF2S23110118	ER-Golgi intermediate compartment 32kDa prote, Dual specificity	growth and immune
38	BICF2S2373811	protein phosphatase1 ER-Golgi intemediate compartment 32kDa prote, Dual specificity	growth and immune
	BICF2P817345	protein phosphatase1 RIKEN Cdna 4930513F16	-
	BICF2G63073656	transforming growth factor, lysophospholopase - like1	growth and immune
	BICF2S23036049	transforming growth factor, lysophospholopase - like1	growth and immune
X	BICF2S2297550	MOCO Sulphurase c-terminal domain containing	formation of institutions
	BICF2S22933746	leucine rich repeats and calponin homology	growth and immune
	BICF2G6306331	glutamine ammonia ligase, odd oz/tenm homolog 1	signal transduction
	BICF2P227876	glutamine ammonia ligase, odd oz/tenm homolog 1	signal transduction
	BICF2G6305980	hypoxantine phosphoribosyltransferse	-
	BICF2P162522	DEADLH box polypeptide2	-
	BICF2P593928	solute carrier family 9	-
	BICF2S23044648	zinc finger protein of the cerellum3	-
	BICF2P1430795	fibroblast growth factor 13 isoform4, ATP binding casseette	growth and immune
	BICF2S23222536	fibroblast growth factor 13 isoform4, ATP binding casseette	growth and immune

### Discussion

We chose Labrador Retriever, as this breed represents the canine HD. From 1974 to 2009 provided by OFA hip dysplasia statistic Bulldog is NO1 in the population studied is 467 dogs and the hip dysplasia 73.2 percentage. According to research provided by OFA hip dysplasia the most many number of breed is Labrador Retriever, Golden Retriever, German Shepherd. Labrador Retrievers are especially studied the highest in 208,931. We performed hip dysplasia of Labrador Retriever SNP 22K chip. canine HD of Labrador Retriever discovered 25 SNPs (Table 1); 25 SNPs located on chromosome 4, 38, X. Until now, research on hip dysplasia SNPs were on chromosome 1, 3, 4, 8, 9, 11, 16, 19, 26, 30, 33 (Budsberg et al., 2006; Marschall and Distl, 2007). In this study, chromosomes 4, 38, X found in the significant SNPs, but a previous study published in the chromosomes of two hip dysplasia 1, 3, 4, 8, 9, 11, 16, 19, 26, 30, 33 overlap in chromosome 4, but in other 38, x chromosome and had found that these SNPs is significant. Hip dysplasia in Labrador Retriever 25 SNPs was found that a variety of roles (Table 2). Significant cell and cell binding, growth and immunity, formation of institutions, signal transduction, divided into four. Also found notable nearby gene is 27 (Table 2). Notable nearby gene is CTNNA3, STOX1, DDX50, COL1A1, USP54, Ndst, PLAU, VDAC2, Sh2d4b, NRG1, BMP 3b precursor, USP6NL, ER-Golgi intermediate compartment 32kDa prote, Dual specificity protein phosphatase1, RIKEN Cdna 4930513F16, TGF, lysophospholopase - like1, MOSC, LRCH, GLUL, ODZ1, HGPT, DEADLH box polypeptide2, SLC9A9, zinc finger protein of the cerellum 3, FGF13 isoform 4, Abc.

Until current research about the human and canine HD association gene is pregnancy associated plasma protein A2 (PAPP-A2) (Jia *et al.*, 2012), cartilage oligomeric matrix protein (COMP), matrilin 3 (MATN3) (Kim *et al.*, 2011), collagen, type IX, alpha 1,2,3 (COL9A·1,2,3) (Dahlqvist et al., 2009; Kim et al., 2011; Nakashima et al., 2005), collagen, type II, alpha 1 (COL2A1) (Kannu et al., 2011), solute carrier family 26 (SLC26) (Hinrichs et al., 2010), fibrillin 2 (FBN2) (Friedenberg *et al.*, 2011), transforming growth factor, beta 1 (TFGβ1), interleukin 6 (IL-6) (Kolundzic *et al.*, 2011), interleukin beta 12 (IL-12β) (Zhou *et al.*, 2010), aspirin (ASPN) (Shi *et al.*, 2011), bone morphogenetic protein (BMP) (Baker-LePain and Lane, 2010), matrix metalloproteinase-1 (MMP-1) (Ray *et al.*, 2005).

These genetic functions of PAPP-A2 is cell differentiation, regulation of cell growth, COMP is cell adhesion, growth plate cartilage development, organ morphlogenesis and skeletal system development. MATN3 functions skeletal system development and COL9A1 has related to cell adhesion, growth plate cartilage development, organ morphogenesis. COL9A2 is skeletal system development, COL9A3 is axon guidance, COL2A1 is axon guidance, skeletal system development, bone development, SLC26 is ossification, FBN2 is bone trabecula formation, positive regulation of bone mineralization, positive regulation of osteoblast differentiation, negative regulation of transforming, growth factor beta receptor signaling, pathway by extracellular sequestering of TGF beta, TGFb1 is cell-cell junction organization, cell death, blood coagulation, aging, IL6 is aging, cell growth, cell redox homeostasis, muscle cell homeostasis, negative regulation of cell proliferation, IL12b is cell cycle arrest, cell migration, natural killer cell activation, negative regulation of smooth muscle cell proliferation, positive regulation of T-cell proliferation, positive regulation of T-helper 1 type immune response, ASPN is bone mineralization,

negative regulation of transforming growth factor growth factor beta receptor signaling pathway, negative regulation of tooth mineralization, BMP is cell differentiation, ossification, positive regulation of cartilage development, proteolysis, skeletal system development, MMP-1 is blood coagulation, collagen catabolic process, proteolysis, leukocyte migration. The common denominator of these genes, cell adhesion, bone formation, skeletal development, is the development of muscle cells.

Until now, the research to the X chromosome is X chromosome associated reports roifman syndrome (Roy, 2011), spondyloepiphyseal dysplasia (Budsberg *et al.*, 2006). Roifman syndrome, spondyloepiphyseal skeletal dysplasia disorders from both, X chromosome disorder that affects the skeletal system to the extent announcement. But this study found X chromosome associated function of genes is formation of institutions, growth and immune, signal transduction. One in the world of genetic disorders caused by inbreeding, and a serious level.

Using genomic information related to the hip displasia single nucleotide polymorphism genotyping. Such as hip dysplasia, a genetic disease is removed, an excellent selection for dogs can be useful as genetic markers. Gene of canine hip dysplasia analysis has not been reported as a genetic marker was not practical. In this study canine genetic improvement hip dysplasia related single nucleotide polymorphism as a genetic marker would be very effective.

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# References

Baker-LePain JC, Lane NE (2010) Relationship between joint shape and the development of osteoarthritis. Curr Opin Rheumatol 22: 538-543.

Budsberg SC, Lenz ME, Thonar EJ (2006) Serum and synovial fluid concentrations of keratan sulfate and hyaluronan in dogs with induced stifle joint osteoarthritis following cranial cruciate ligament transection. Am J Vet Res 67: 429-432.

Dahlqvist J, Orlen H, Matsson H, Dahl N, Lonnerholm T, Gustavson KH (2009) Multiple epiphyseal dysplasia. Acta Orthop 80: 711-715.

Friedenberg SG, Zhu L, Zhang Z, Foels W, Schweitzer PA, Wang W, Fisher PJ, Dykes NL, Corey E, Vernier-Singer M, Jung SW, Sheng X, Hunter LS, McDonough SP, Lust G, Bliss SP, Krotscheck U, Gunn TM, Todhunter RJ (2011) Evaluation of a fibrillin 2 gene haplotype associated with hip dysplasia and incipient osteoarthritis in dogs. Am J Vet Res 72: 530-540.

Goldberg RL, Rubin AS (1989) Serum hyaluronate as a marker for disease severity in the Lactobacillus casei model of arthritis in the rat. J Rheumatol 16: 92-96.

Hinrichs T, Superti-Furga A, Scheiderer WD, Bonafe L, Brenner RE, Mattes T (2010) Recessive multiple epiphyseal dysplasia (rMED) with homozygosity for C653S mutation in the DTDST gene--phenotype, molecular diagnosis and surgical treatment of habitual dislocation of multilayered patella: case report. BMC

Musculoskelet Disord 11: 110.

Jia J, Li L, Zhao Q, Zhang L, Ru J, Liu X, Li Q, Shi L (2012) Association of a single nucleotide polymorphism in pregnancy-associated plasma protein-A2 with developmental dysplasia of the hip: a case-control study. Osteoarthritis Cartilage 20: 60-63.

Kannu P, Irving M, Aftimos S, Savarirayan R (2011) Two novel COL2A1 mutations associated with a Legg-Calve-Perthes disease-like presentation. Clin Orthop Relat Res 469: 1785-1790.

Kim OH, Park H, Seong MW, Cho TJ, Nishimura G, Superti-Furga A, Unger S, Ikegawa S, Choi IH, Song HR, Kim HW, Yoo WJ, Shim JS, Chung CY, Oh CW, Jeong C, Song KS, Seo SG, Cho SI, Yeo IK, Kim SY, Park S, Park SS (2011) Revisit of multiple epiphyseal dysplasia: ethnic difference in genotypes and comparison of radiographic features linked to the COMP and MATN3 genes. Am J Med Genet A 155A: 2669-2680.

Kolundzic R, Trkulja V, Mikolaucic M, Kolundzic MJ, Pavelic SK, Pavelic K (2011) Association of interleukin-6 and transforming growth factor-beta1 gene polymorphisms with developmental hip dysplasia and severe adult hip osteoarthritis: a preliminary study. Cytokine 54: 125-128.

Marschall Y, Distl O (2007) Mapping quantitative trait loci for canine hip dysplasia in German Shepherd dogs. Mamm Genome 18: 861-870.

Nakashima E, Kitoh H, Maeda K, Haga N, Kosaki R, Mabuchi A, Nishimura G, Ohashi H, Ikegawa S (2005) Novel COL9A3 mutation in a family with multiple epiphyseal dysplasia. Am J Med Genet A 132A: 181-184.

Nganvongpanit K, Itthiarbha A, Ong-Chai S, Kongtawelert P (2008) Evaluation of serum chondroitin sulfate and hyaluronan: biomarkers for osteoarthritis in canine hip dysplasia. J Vet Sci 9: 317-325.

Quignon P, Herbin L, Cadieu E, Kirkness EF, Hedan B, Mosher DS, Galibert F, Andre C, Ostrander EA, Hitte C (2007) Canine population structure: assessment and impact of intra-breed stratification on SNP-based association studies. PLoS One 2: e1324.

Ray A, Shakya A, Ray BK (2005) Inflammation-responsive transcription factors SAF-1 and c-Jun/c-Fos promote canine MMP-1 gene expression. Biochim Biophys Acta 1732: 53-61.

Roy DR (2011) Spectrum of intra-articular findings of the acute and subacute painful hip with multiple epiphyseal dysplasia/spondyloepiphyseal dysplasia. J Pediatr Orthop B 20: 284-286.

Shi D, Dai J, Zhu P, Qin J, Zhu L, Zhu H, Zhao B, Qiu X, Xu Z, Chen D, Yi L, Ikegawa S, Jiang Q (2011) Association of the D repeat polymorphism in the ASPN gene with developmental dysplasia of the hip: a case-control study in Han Chinese. Arthritis Res Ther 13: R27.

Zhou Z, Sheng X, Zhang Z, Zhao K, Zhu L, Guo G, Friedenberg SG, Hunter LS, Vandenberg-Foels WS, Hornbuckle WE, Krotscheck U, Corey E, Moise NS, Dykes NL, Li J, Xu S, Du L, Wang Y, Sandler J, Acland GM, Lust G, Todhunter RJ (2010) Differential genetic regulation of canine hip dysplasia and osteoarthritis. PLoS One 5: e13219.